A new example of a steroid-amino acid hybrid: construction of constrained nine membered D-ring steroids[†]‡

Shagufta and Gautam Panda*

Received 6th November 2006, Accepted 10th November 2006 First published as an Advance Article on the web 4th December 2006 DOI: 10.1039/b616155c

A new archetype of steroid–amino acid hybrids containing a nine membered D-ring with hetero atoms has been synthesized for the first time from estrone and amino acids by using Yamaguchi coupling reactions.

Introduction

Nature remains the unending source for biologically important natural products of molecular diversity and complexity. These naturally occurring molecules, which are essentially derived from particular or combined biosynthetic pathways, possess novel physical, chemical and biological properties.¹ The human quest to acquire novel molecules with interesting properties has generated an idea of architecting an inexhaustible reservoir of novel synthetic endeavours by rationally combining two or more different classes of natural products.² During the past two decades, the design and synthesis of such molecular structures have received much attention and these have been referred to as 'hybrid molecules'.

Hybrids or conjugates between hydrophobic steroids and hydrophilic amino acids or peptides, i.e. peptidyl steroids, have been reported to play important roles to enhance oral antiarrhythmic activity, to promote delivering prodrugs to specific target tissues and to achieve 'permissive action'.3-5 Several peptidyl steroids such as polymastiamide A, a tyrosine conjugated steroid analog isolated from the Norwegian marine sponge Polymastia boletiformis exhibiting antimicrobial activity and bufetoxin, a 3-arginyl derived steroid product isolated from the Chinese hoptoad have been reported.6 Moreover, cholyl glycine or cholyl taurine which exists in the bile of animals and contains a glycyl or a taurinyl group at the 17-position of the steroid, has led to the development of several steroid-amino acid conjugates for estrogen dependent biological activity,^{7 14}C-cholyl-glycine breath test,⁸ and as antidepressant agents.9 To date, it is mainly peptidyl steroids of the androstane and estrone series comprising amino acids or peptides attached with the unchanged nucleus of steroids through an amide or ester bond, that have been synthesized and examined. There is no reported example of a chiral amino acid incorporated D-ring modified peptidyl steroid.

Results and discussion

Here we describe the design and synthesis of a new archetype of 'steroid-amino acid hybrid' 1 with amino acids incorporated

Medicinal and Process Chemistry Division, Central Drug Research Institute, Chattar Manzil Palace, Lucknow 226001, UP, India. E-mail: gautam.panda@gmail.com

[‡] Electronic supplementary information (ESI) available: ¹H and ¹³C NMR spectra of compounds **3–17** and ¹H-¹H COSY, HSQC and HMBC spectra of **3b**. See DOI: 10.1039/b616155c

onto the D-ring of steroid **2** (Fig. 1). The projected molecule **1**, having a constrained nine membered D-ring with hetero atoms on the steroidal framework, provides a highly efficient access to novel molecular hybrids. To the best of our knowledge, only one example of a nine membered D-ring on a steroid using Heck coupling is reported.¹⁰ Intramolecular macrolactonization on a steroidal framework to furnish a nine membered D-ring has not been reported so far.

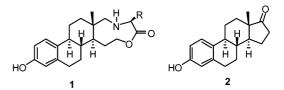
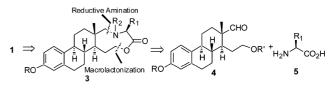


Fig. 1 Structures of projected molecule 1 and estrone 2.

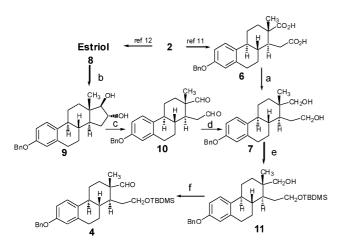
Retrosynthetic analysis revealed that projected molecule **1** could be accessed from **3** which could be obtained from building blocks *seco*-estrone aldehyde **4** and amino acid **5** through utilization of reductive amination and macrolactonization (Scheme 1).



Scheme 1 Retrosynthetic analysis of target molecule.

The synthesis of *seco*-estrone aldehyde **4** was accomplished from readily available estrone **2** (Scheme 2). Alkylation of estrone **2** with benzyl bromide– K_2CO_3 followed by base and an iodine mediated haloform reaction gave marrianolic acid benzyl ether **6** (75%).¹¹ LAH reduction of **6** led to diol **7** (35%). The overall yield of **7** from **2** was very poor (26%). It was therefore decided to develop an alternative strategy for large scale access to **7**. Towards this objective, estriol **8** was initially synthesized from **2** following literature procedures.¹² The phenolic hydroxy of **8** was benzylated to give **9** (95%). The vicinal hydroxyl groups of estriol 3-benzyl ether **9** were subjected to periodate cleavage (NaIO₄) to deliver the dialdehyde **10** which was subjected to LAH reduction to furnish **7** with 62% overall yield. Further, the C₁₆ hydroxyl group of diol **7** was selectively protected as TBDMS derivative **11** (67%) by treatment of alcohol **7** with 1 : 1 equivalents of TBDMSCI

[†] CDRI Communication number 6739.



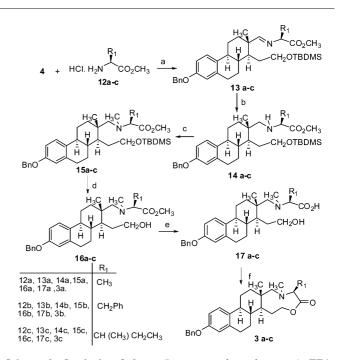
Scheme 2 Synthesis of key intermediate D-*seco* estrone aldehyde 4. *Reagents and conditions:* a) LAH, THF, 0-5 °C, 1 h, 35%; b) benzyl bromide, K₂CO₃, acetone, reflux, 8 h, 95%; c) NaIO₄, methanol–H₂O (4 : 1), 0 °C, 24 h; d) LAH, THF, 0-5 °C, 1 h, 81% (two steps); e) TBDMSCl, imidazole, DCM, 0 °C–rt, 6 h, 67%; f) TPAP, NMO, MS 4 Å, DCM, rt, 1 h, 82%.

and imidazole. TPAP mediated oxidation of the primary hydroxyl group in **11** led to the key aldehyde **4** in 82% yield. The aldehyde **4** was prepared on a large scale for the execution of reductive amination with chiral amino acid ester.

Reductive amination of **4** with amino acid methyl ester hydrochloride¹³ **12a–c** in the presence of NaBH₃CN and acetic acid was performed to afford amine **14a** (30%), **14b** (34%) and **14c** (31%). Yield of the amine **14a–c** was improved (80%, 78% and 73% respectively) by performing the reaction in two steps through initial formation of imine **13a–c** followed by reduction with NaBH₄. It was decided to convert the secondary amine to a tertiary one to avoid unnecessary reactions in the next steps. Treatment of amine **14a–c** with MeI–K₂CO₃ in acetone furnished the fully protected product **15a** (97%), **15b** (95%) and **15c** (93%). Upon exposure to AcOH–H₂O–THF (3 : 1 : 1) at 50–60 °C, deprotection of TBDMS ether occurred to afford the alcohol **16a** (66%), **16b** (63%) and **16c** (70%).

Finally the methyl ester of **16a–c** was hydrolyzed in the presence of 1 N NaOH–dioxane to furnish the hydroxy acid **17a** (70%), **17b** (67%) and **17c** (72%). The hydroxy acid was then subjected to macrolactonization using the Yamaguchi method¹⁴ to furnish the corresponding lactone **3a** (53%), **3b** (63%) and **3c** (42%) which can afford the target molecule **1** on deprotection of a benzyl group, Scheme 3. The structure of steroid–amino acid hybrid **3a–c** was secured from incisive analysis of ¹H, ¹³C NMR, ¹H-¹H COSY, HSQC and HMBC spectra.

In conclusion, we have demonstrated a new prototype of steroid–amino acid hybrid 1 from easily available estrone 2 and amino acids *i.e.* alanine, phenyl alanine and isoleucine through a series of simple and efficient steps involving NaIO₄ cleavage of estriol, reductive amination and Yamaguchi coupling reactions. A simple and convenient approach to access chiral amino acid incorporated D-ring modified steroid–amino acid hybrids (peptidyl steroids) is described. In the process, we have synthesized several important molecules such as diol, D-*seco* estrone aldehyde and and D-*seco* estrone amino acid conjugates which can be used as advanced intermediates for accessing new steroid derivatives.



Scheme 3 Synthesis of 3a–c. *Reagents and conditions:* a) TEA, methanol–THF (4 : 1), reflux–rt, 12 h; b) NaBH₄, methanol, 0–5 °C, 1 h, 14a = 80%, 14b = 78% and 14c = 73% for two steps; c) MeI, K₂CO₃, acetone, rt, 24 h, 15a = 97%, 15b = 95% and 15c = 93%; d) AcOH–H₂O–THF (3 : 1 : 1), 60 °C, 2 h, 16a = 66%, 16b = 63% and 16c = 70%; c) 1 N NaOH, dioxane, 60 °C, 1 h, 17a = 70%, 17b = 67% and 17c = 72%; f) 2,4,6-Cl₃C₆H₂COCl, Et₃N, THF, rt, then DMAP, PhMe, reflux, 3a = 53%, 3b = 63% and 3c = 42%.

Additionally, the acid functionality of **17a–c** can be further coupled with other amino acids to elaborate macrocyclic steroid–amino acid hybrids for interesting biological functions. Efforts in this direction are in progress.

Experimental

General

All the reactions were monitored by thin layer chromatography over silica gel coated TLC plates. The spots on TLC were visualized by warming ceric sulfate (2% CeSO₄ in 2 N H₂SO₄) sprayed plates on a hot plate or in an oven at about 100 °C. Silica gel 60-120 mesh was used for column chromatography. Melting points were recorded on an electrically heated apparatus and are uncorrected. IR spectra were recorded on a Perkin Elmer 881 or FT IR 820/PC instrument and values are expressed in cm⁻¹. Electron impact mass spectra were recorded on a JEOL (Japan)/D-300 instrument and FAM mass spectra were recorded on a JEOL SX 102/DA-6000 mass spectrometer using Argon/Xenon (6 KV, 10 MA) as the FAB gas. ¹H and ¹³C NMR spectra were recorded on a Bruker Avance DPX 200 MHz instrument using TMS as an internal reference. Chemical shift values are expressed in δ ppm. Specific rotation was determined with a Rudolph Autopol IIIrd polarimeter at 28 °C. Elementary analysis was carried out on a Carlo ERBA-1108 analyzer. Commercially available grades of organic solvents of adequate purity were used in many reactions. Acetone was refluxed with KMnO₄ for 4 h, after that it was distilled and stored in a bottle containing dried K₂CO₃. Benzene was refluxed with freshly

cut and dried sodium metal pieces pressed in 3 Å sieves for 4–6 h. It was distilled and stored in a dry bottle. Tetrahydrofuran was dried initially over calcium sulfate and then refluxed over lithium aluminium hydride. Peroxide was removed by passage through a column of alumina and distilled and stored over molecular sieves 3 Å.

Experimental procedure

Diol 7. To a stirred solution of estriol 3-benzyl ether **9** (2.62 g, 6.93 mmol) in methanol (40 ml) at 0 °C was added a solution of NaIO₄ (2.22 g, 10.40 mmol) in water (10 ml). After stirring at 0 °C for 24 h, the methanol was evaporated *in vacuo* at low temperature. The aqueous solution was extracted with CH₂Cl₂ twice. The combined organic layer was washed with water, brine, dried over Na₂SO₄ and concentrated under vacuum to yield dialdehyde **10** (3.0 g) as a yellow viscous oil which was used as such for the next step.

Dialdehyde 10 (3.0 g) in THF (5 ml) was added dropwise to a stirred solution of LAH (0.45 g, 11.97 mmol) in 40 ml of THF at 0 °C. The resulting solution was stirred at rt for 1 h. After completion (monitored by TLC), the reaction was quenched with ethyl acetate followed by water at 0 °C. The aqueous layer was extracted twice with ethyl acetate. The combined organic layer was washed with brine, dried over anhydrous Na2SO4 and concentrated under vacuum. The residue was chromatographed over silica gel and elution with 40% ethyl acetate in hexane ($R_f = 0.5$) furnished diol 7 (2.12 g, 81%) as white solid, mp 135 °C. IR (KBr): 3336, 2920, 1606, 1255, 1035, 740 cm⁻¹. ¹H NMR (200 MHz, CDCl₃): δ 7.44-7.20 (m, 6H, -O-CH₂-C₆H₅ and C-1-H), 6.78 (dd, 1H, $J_1 = 8.6, J_2 = 2.4, C-2-H), 6.70 (d, 1H, J = 2.4, C-4-H), 5.02$ (s, 2H, -O-CH₂-C₆H₅), 4.00-3.85 (m, 1H, C-16-H), 3.67 (d, 1H, J = 11.7, C-17-H, 3.61–3.46 (m, 1H, C-16-H), 3.09 (d, 1H, J =11.7, C-17-H), 2.85–2.83 (m, 2H, C-6-H₂), 2.33–2.26 (m, 2H), 2.01–1.95 (m, 2H), 1.75–1.28 (m, 7H), 0.69 (s, 3H, C-18-H₃). ¹³C NMR (50 MHz, CDCl₃): δ 157.1 (C-3), 138.2 (C-5), 137.7, 133.4 (C-10), 128.9, 128.2, 127.9, 127.0 (C-1), 114.8 (C-4), 112.9 (C-2), 70.5 (C-17), 70.4 (-O-CH2-C6H5), 64.3 (C-16), 43.9, 42.2, 41.2, 38.9 (C-13), 35.7, 31.0, 30.3 (C-6), 27.8 (C-12), 27.0 (C-11), 16.4 (C-18). MASS (FAB): m/z (%): 380 (100, [M⁺]), 363 (40, [M⁺-OH)]). Anal. Calcd for (C₂₅H₃₂O₃): C, 78.91; H, 8.48%. Found: C, 79.17; H, 8.60%.

TBDMS derivative 11. To a solution of diol 7 (2 g, 5.26 mmol) in CH₂Cl₂ (20 ml) at 0 °C was added imidazole (0.36 g, 5.26 mmol) followed by TBDMSCl (0.79 g, 5.26 mmol). The mixture was stirred at rt overnight. The reaction was quenched with H₂O and diluted with CH₂Cl₂. The aqueous layer was extracted with CH₂Cl₂ twice. The combined organic layers were dried over Na₂SO₄ and concentrated under vacuum. The residue was chromatographed over silica gel and elution with 10% ethyl acetate in hexane ($R_{\rm f} =$ 0.5) furnished 11 (1.75 g, 67%) as a white solid, mp 77 $^{\circ}$ C with recovery of starting material 7 (0.592 g, 30%). IR (KBr): 3473, 2927, 1500, 1463, 1253, 1046, 836 cm⁻¹. ¹H NMR (200 MHz, CDCl₃): δ 7.28–7.22 (m, 5H, –O–CH₂–C₆H₅), 7.12 (d, 1H, J = 8.6, C-1-*H*), 6.69 (dd, 1H, $J_1 = 8.6$, $J_2 = 2.4$, C-2-*H*), 6.59 (d, 1H, J = 2.4, C-4-H, 4.97 (s, 2H, $-O-CH_2-C_6H_5$), 3.85–3.69 (m, 1H, C-16-*H*), 3.55 (d, 1H, J = 12, C-17-*H*), 3.43–3.32 (m, 1H, C-16-*H*), 2.95 (d, 1H, J = 12, C-17-H), 2.73–2.72 (m, 2H, C-6-H₂), 2.32–

2.09 (m, 2H), 1.95–1.91 (m, 9H), 0.81 {s, 9H, $-C(CH_3)_3$ }, 0.56 (s, 3H, C-18- H_3), 0.04 {s, 6H, $-Si(CH_3)_2-$ }. ¹³C NMR (50 MHz, CDCl₃): δ 157.1 (C-3), 138.1 (C-5), 137.7, 133.4 (C-10), 128.9, 128.2, 127.8, 127.1 (C-1), 114.9 (C-4), 112.9 (C-2), 70.9 (C-17), 70.3 ($-O-CH_2-C_6H_5$), 65.8 (C-16), 44.0, 42.3, 41.0, 38.9 (C-13), 35.8, 31.0, 30.8 (C-6), 27.8 (C-12), 27.1(C-11), 26.5 { $-C(CH_3)_3$ }, 19.1{ $-C(CH_3)_3$ }, 16.3 (C-18), -4.9 {Si(CH_3)_2-}. MASS (FAB): m/z (%): 495 (40, [M⁺ + H]), 363 (70, [M⁺–OH,-TBDMS]). Anal. Calcd for (C₃₁H₄₆O₃Si): C, 75.25; H, 9.37%. Found: C, 75.42; H, 9.64%.

Aldehyde 4. To a solution of 11 (1 g, 2.02 mmol) in CH₂Cl₂ (25 ml) were added molecular sieves (4 Å) followed by NMMO (0.41 g, 3.04 mmol). After stirring the mixture for 10 min, TPAP (0.07 g, 0.20 mmol) was added and it was stirred at rt for 2 h. After completion (monitored by TLC), the reaction mixture was diluted with CH2Cl2 and was washed with sodium sulfite, brine, saturated $CuSO_4$ solution, dried over Na_2SO_4 and concentrated *in vacuo*. The residue was chromatographed over silica gel and elution with 10% ethyl acetate in hexane ($R_f = 0.6$) furnished 4 (0.82 g, 82%) as white solid, mp 80 °C. IR (KBr): 3417, 2945, 1724, 1598, 1359, 1093, 837, 771 cm⁻¹. ¹H NMR (200 MHz, CDCl₃): δ 9.37 (s, 1H, C-17-*H*), 7.36–7.29 (m, 5H, –O–CH₂–C₆ H_5), 7.16 (d, 1H, J = 8.6, C-1-*H*), 6.71 (dd, 1H, $J_1 = 8.6$, $J_2 = 2.4$, C-2-*H*), 6.67 (d, 1H, J = 2.4, C-4-H), 4.98 (s, 2H, -O- CH_2 - C_6H_5), 3.61–3.39 (m, 2H, C-16-H₂), 2.72-2.88 (m, 2H, C-6-H₂), 2.35-2.22 (m, 2H), 2.15-2.02 (m, 1H), 1.71-1.15 (m, 8H), 1.00 (s, 3H, C-18-H₃), 0.85 {s, 9H, $-C(CH_3)_3$, 0.00 {s, 6H, $-Si(CH_3)_2$ -}. ¹³C NMR (50 MHz, CDCl₃): δ 207.0 (C-17), 157.3 (C-3), 138.2 (C-5), 137.6, 132.5 (C-10), 128.9, 128.3, 127.8, 126.8 (C-1), 114.9 (C-4), 113.0 (C-2), 70.3 (-O-CH₂-C₆H₅), 63.4 (C-16), 51.1 (C-13), 43.6, 41.0, 40.6, 34.1, 33.0, 30.7 (C-6), 27.4 (C-12), 26.4 {-C(CH₃)₃}, 25.8 (C-11), $18.7\{-C(CH_3)_3\}, 13.4 (C-18), -4.8 \{Si(CH_3)_2-\}. MASS (FAB):$ m/z (%): 492 (60, [M⁺]), 491 (80, [M⁺-H]). Anal. Calcd for (C₃₁H₄₄O₃Si): C, 75.56; H, 9.00%. Found: C, 75.76; H, 9.29%.

Amine 14a. To a solution of alanine methyl ester hydrochloride 12a (0.16 g, 1.13 mmol) in methanol (12 ml) was added triethyl amine (0.21 ml, 1.50 mmol). After stirring the mixture for 5 min, a solution of aldehyde 4(0.37 g, 0.75 mmol) in THF (3 ml) was added and the reaction mixture was refluxed for 2 h. After stirring at rt overnight, the solvent was removed and the residue was dissolved in ethyl acetate. The undissolved material was filtered off and the filtrate was concentrated to afford the crude imine 13a (0.4 g).

To a solution of crude imine 13a (0.4 g) in methanol (15 ml), $NaBH_4$ (0.04 gm, 1.04 mmol) was added in a small portion at 0 °C. After 15 min, the solvent was removed under vacuum and the residue was passed through silica gel and elution with 10% ethyl acetate in hexane ($R_f = 0.4$) furnished **14a** (0.35 g, 80%) as a white solid, mp 72 °C, [a]²⁰_D +17 (c 2, MeOH). IR (KBr): 3570, 2935, 1737, 1598, 1463, 1251, 1093, 840, 775 cm⁻¹. ¹H NMR (200 MHz, CDCl₃): δ 7.39–7.02 (m, 6H, –O–CH₂–C₆H₅ and C-1-H), 6.61 (dd, $1H, J_1 = 8.6, J_2 = 2.4, C-2-H), 6.58 (d, 1H, J = 2.4, C-4-H), 4.94$ (s, 2H, -O-CH₂-C₆H₅), 3.65 (s, 3H, -CO₂CH₃), 3.63-3.39 (m, 2H, C-16-H₂), 3.26–3.06 (m, 1H, –CH–CH₃), 2.80–2.62 (m, 2H, C-6-H₂), 2.29 (s, 2H, C-17-H₂), 2.28–2.01 (m, 3H), 1.65–1.47 (m, 4H), 1.40–1.09 (m, 4H), 1.22 (d, 3H, J = 7, –CH–CH₃), 0.84 {s, 9H, $-C(CH_3)_3$, 0.72 (s, 3H, C-18- H_3), 0.00 {s, 6H, $-Si(CH_3)_2$ -}. ¹³C NMR (50 MHz, CDCl₃): δ 177.1 (-CO₂CH₃), 157.1 (C-3), 138.4 (C-5), 137.9, 133.5 (C-10), 128.9, 128.2, 127.8, 126.9 (C- 1), 114.8 (C-4), 112.8 (C-2), 70.3 ($-O-CH_2-C_6H_3$), 64.8 (C-16), 58.4 (C-17), 57.8 ($-CH-CH_3$), 52.0 ($-CO_2CH_3$), 43.9, 42.4, 42.1, 37.9 (C-13), 36.9, 32.8, 31.0 (C-6), 28.0 (C-12), 26.9 (C-11), 26.5 { $-C(CH_3)_3$ }, 19.4 { $-C(CH_3)_3$ }, 18.9 ($-CH-CH_3$), 18.1 (C-18), -4.7 {Si($CH_3)_2$ -}. MASS (FAB): m/z (%): 580 (100, M⁺). Anal. Calcd for ($C_{35}H_{33}NO_4Si$): C, 72.49; H, 9.21; N, 2.42%. Found: C, 72.71; H, 9.39; N, 2.51%.

Amine 14b. As described for 14a, phenylalanine methyl ester hydrochloride 12b (0.20 g, 0.91 mmol) in methanol (15 ml), triethyl amine (0.17 ml, 1.22 mmol), aldehyde 4 (0.3 g, 0.61 mmol) in THF (3 ml) furnished imine 13b (0.33 g).

13b (0.33 g, 0.505 mmol) in methanol (15 ml), NaBH₄ (0.03 g, 0.76 mmol) furnished 14b (0.31 g, 78%) as a viscous oil, $R_{\rm f}$ = 0.45 (10% ethyl acetate in hexane), $[a]_{D}^{20}$ +27 (c 2, MeOH). IR (Neat): 3427, 2931, 1737, 1598, 1355, 1091, 839, 773 cm⁻¹. ¹H NMR (200 MHz, CDCl₃): δ 7.33–7.11 (m, 11H, –O–CH₂–C₆H₅, $-CH-CH_2-C_6H_5$ and C-1-H), 6.58 (dd, 1H, $J_1 = 8.6, J_2 = 2.4, C-1$ 2-*H*), 6.65 (d, 1H, J = 2.4, C-4-*H*), 4.94 (s, 2H, $-O-CH_2-C_6H_5$), 3.56 (s, 3H, $-CO_2CH_3$), 3.46–3.34 (m, 3H, C-16- H_2 and $-CH_2$ $CH_2-C_6H_5$, 2.85 (d, 2H, J = 6.8, $-CH-CH_2-C_6H_5$), 2.83–2.77 (m, 2H, C-6-H₂), 2.44–2.34 (m, 2H, C-17-H₂), 2.26–2.09 (m, 3H), 2.02 $(bs, 1H, -NH-), 1.59-1.19 (m, 8H), 0.84 \{s, 9H, -C-(CH_3)_3\}, 0.68$ (s, 3H, C-18- H_3), 0.00 {s, 6H, $-Si(CH_3)_2-$ }. ¹³C NMR (50 MHz, CDCl₃): δ 175.8 (-CO₂CH₃), 157.1 (C-3), 138.4 (C-5), 138.0, 137.7, 133.5 (C-10), 129.7, 128.9, 128.7, 128.2, 127.8, 127.0 (C-1), 114.8 (C-4), 112.8 (C-2), 70.3 (-O-CH₂-C₆H₅), 64.7 (C-16), 64.5 (-CH-CH₂-C₆H₅), 58.5 (C-17), 51.9 (-CO₂CH₃), 44.0, 42.4, 42.2, 40.4 (-CH-CH₂-C₆H₅), 38.2 (C-13), 36.8, 32.9, 31.0 (C-6), 28.0 (C-12), 26.9 (C-11), 26.4 {-C(CH₃)₃}, 18.8 {-C(CH₃)₃}, 18.1 (C-18), -4.6 ${Si(CH_3)_2-}$. MASS (FAB): m/z (%): 656 (100, M⁺ + H). Anal. Calcd for (C₄₁H₅₇NO₄Si): C, 75.07; H, 8.76; N, 2.14%. Found: C, 75.18; H, 8.88; N, 2.05%.

Amine 14c. As described for 14a, isoleucine methyl ester hydrochloride 12c (0.17 g, 0.91 mmol) in methanol (15 ml), triethyl amine (0.17 ml, 1.22 mmol), aldehyde 4 (0.3 g, 0.61 mmol) in THF (3 ml) furnished imine 13c (0.35 g).

13c (0.35 g, 0.57 mmol) in methanol (15 ml), NaBH₄ (0.03 g, 0.85 mmol) furnished 14c (0.28 g, 73%) as a viscous oil, $R_{\rm f}$ = 0.5 (10% ethyl acetate in hexane), $[a]_{D}^{20}$ +19.5 (c 2, MeOH). IR (Neat): 3447, 2930, 1733, 1609, 1500, 1463, 1382, 1252, 1091, 837, 759 cm⁻¹. ¹H NMR (200 MHz, CDCl₃): δ 7.36–7.21 (m, 5H, $-O-CH_2-C_6H_5$, 7.19–7.03 (d, 1H, J = 8.6, C-1-H), 6.65 (dd, 1H, $J_1 = 8.6, J_2 = 2.4, C-2-H), 6.62 (d, 1H, J = 2.4, C-4-H), 4.92 (s, J_1 = 2.4, C-4-H), 4.92 (s, J_2 = 2.4, C-4-$ 2H, $-O-CH_2-C_6H_5$), 3.63 (s, 3H, $-CO_2CH_3$), 3.52–3.35 (m, 2H, C-16-H₂), 3.52–3.35 {m, 1H, -CH-CH-(CH₃)-CH₂-CH₃}, 2.80-2.68 (m, 2H, C-6-H₂), 2.35–1.89 (m, 5H), 1.62–1.14 (m, 15H), 0.83 {bs, 12H, -C(CH₃)₃}, 0.69 (s, 3H, C-18-H₃), 0.00 {s, 6H, $-Si(CH_3)_2$ -}. ¹³C NMR (50 MHz, CDCl₃): δ 176.5 ($-CO_2CH_3$), 157.1 (C-3), 139.2 (C-5), 138.4, 133.6 (C-10), 128.9, 128.2, 127.8, 126.9 (C-1), 114.9 (C-4), 112.8 (C-2), 70.3 (-O-CH₂-C₆H₅), 67.9 {-CH-CH-(CH₃)-CH₂-CH₃}, 64.8 (C-16), 59.0 (C-17), 51.6 (-CO₂CH₃), 44.0, 42.4, 42.2, 39.0 {-CH-CH-(CH₃)-CH₂-CH₃}, 38.2 (C-13), 37.0, 33.0, 31.1 (C-6), 28.1 (C-12), 26.9 (C-11), 26.5 $\{-C(CH_3)_3\}$, 26.2 $\{-CH-CH-(CH_3)-CH_2-CH_3\}$, 18.8 $\{-C(CH_3)_3\}, 18.3 (C-18), 16.2 \{-CH-CH-(CH_3)-CH_2-CH_3\},\$ 12.0 $\{-CH-CH-(CH_3)-CH_2-CH_3\}, -4.6 \{Si(CH_3)_2-\}$. MASS (FAB): m/z (%): 621 (100, M⁺). Anal. Calcd for (C₃₈H₅₉NO₄Si): C, 73.38; H, 9.56; N, 2.25%. Found: C, 73.45; H, 9.25; N, 2.12%.

N-Methyl amine derivative 15a. A mixture of 14a (0.2 g, 0.35 mmol), anhydrous K₂CO₃ (0.24 g, 1.72 mmol), methyl iodide (0.04 ml, 0.69 mmol) and dry acetone (15 ml) was stirred at room temperature for 24 h. K₂CO₃ was filtered off and the acetone was removed in vacuo. The residue was diluted with water and extracted with ethyl acetate. The organic layer was washed with water, brine, dried over anhydrous Na₂SO₄ and concentrated under vacuum. The residue was chromatographed over silica gel and elution with 5% ethyl acetate in hexane ($R_f = 0.45$) furnished **15a** (0.20 g, 97%) as a white semi solid, $[a]_{D}^{20}$ +39.5 (c 2, MeOH). IR (KBr): 3460, 2932, 1735, 1609, 1500, 1461, 1248, 1091, 772 $\rm cm^{-1}.$ $^1\rm H~NMR$ $(200 \text{ MHz}, \text{CDCl}_3)$: δ 7.30–7.17 (m, 5H, –O–CH₂–C₆H₅), 7.05 (d, 1H, J = 8.6, C-1-H), 6.69 (dd, 1H, $J_1 = 8.6$, $J_2 = 2.4$, C-2-H), 6.57 (d, 1H, J = 2.4, C-4-H), 4.93 (s, 2H, $-O-CH_2-C_6H_5$), 3.60 (s, 3H, -CO₂CH₃), 3.59–3.18 (m, 3H, C-16-H₂ and -CH-CH₃), 2.81–2.67 (m, 2H, C-6- H_2), 2.55 (d, 1H, J = 11.8, C-17-H), 2.31 (s, $3H, -N-CH_3$, 2.29 (d, 1H, J = 11.8, C-17-H), 2.29–1.99 (m, 3H), $1.81-1.26 \text{ (m, 8H)}, 1.21 \text{ (d, 3H, } J = 7.1, -CH-CH_3), 0.84 \text{ {s, 9H}},$ $-C(CH_3)_3$, 0.77 (s, 3H, C-18-H₃), 0.01 {s, 6H, $-Si(CH_3)_2$ -}.¹³C NMR (50 MHz, CDCl₃): δ 174.7 (-CO₂CH₃), 157.1 (C-3), 138.3 (C-5), 137.7, 133.5 (C-10), 128.9, 128.2, 127.8, 127.0 (C-1), 114.8 (C-4), 112.8 (C-2), 70.3 (-O-CH2-C6H5), 67.8 (C-17), 64.8 (C-16), 64.0 (-CH-CH₃), 51.4 (-CO₂CH₃), 44.6, 44.1, 42.2, 40.7 (-N-CH₃), 40.2 (C-13), 37.9, 33.2, 31.0 (C-6), 28.0 (C-12), 26.9 (C-11), $26.5 \{-C(CH_3)_3\}, 18.9 \{-C(CH_3)_3\}, 16.7 (-CH-CH_3), 16.0 (C-18),$ $-4.6 {Si(CH_3)_2-}$. MASS (FAB): m/z (%): 594 (100, M⁺). Anal. Calcd for (C₃₆H₅₅NO₄Si): C, 72.80; H, 9.33; N, 2.36%. Found: C, 72.96; H, 9.49; N, 2.45%.

N-Methyl amine derivative 15b. As described for 15a, amine 14b (0.22 g, 0.34 mmol), K₂CO₃ (0.23 g, 1.68 mmol), methyl iodide (0.04 ml, 0.67 mmol) in acetone (15 ml) furnished **15b** (0.21 g, 95%) as a white semi-solid, $R_{\rm f} = 0.5$ (5% ethyl acetate in hexane), $[a]_{\rm D}^{20}$ +12.5 (c 2, MeOH). IR (KBr): 3375, 2935, 1738, 1598, 1352, 1245, 1089, 771 cm⁻¹. ¹H NMR (200 MHz, CDCl₃): δ 7.32–7.08 (m, 11H, $-O-CH_2-C_6H_5$, $-CH-CH_2-C_6H_5$ and C-1-H), 6.68 (dd, 1H, $J_1 =$ $8.6, J_2 = 2.4, C-2-H), 6.61 (d, 1H, J = 2.4, C-4-H), 4.93 (s, 2H, -O CH_2-C_6H_5$, 3.58 (s, 3H, $-CO_2CH_3$), 3.48–3.23 (m, 3H, C-16- H_2 and -CH-CH₂-C₆H₅), 2.97-2.71 (m, 2H, -CH-CH₂-C₆H₅), 2.69-2.52 (m, 2H, C-6-H₂), 2.38–2.24 (m, 5H, C-17-H₂ and –N–CH₃), 2.09-1.14 (m, 11H), 0.83 {s, 9H, -C-(CH₃)₃}, 0.63 (s, 3H, C-18- H_3), 0.00 {s, 6H, -Si(C H_3)₂-}.¹³C NMR (50 MHz, CDCl₃): δ 173.3 (-CO₂CH₃), 157.1 (C-3), 139.3 (C-5), 138.3, 137.8, 133.6 (C-10), 129.9, 128.9, 128.6, 128.2, 127.8, 127.0 (C-1), 126.7, 114.7 (C-4), 112.8 (C-2), 71.0 (-CH-CH₂-C₆H₅), 70.3 (-O-CH₂-C₆H₅), 68.2 (C-17), 64.7 (C-16), 51.4 (-CO₂CH₃), 43.7, 43.5, 42.3, 40.5 (-N-CH₃), 40.4 (C-13), 37.4, 36.5 (-CH-CH₂-C₆H₅), 33.2, 31.0 (C-6), 27.9 (C-12), 26.9 (C-11), 26.5 $\{-C(CH_3)_3\}$, 18.9 $\{-C(CH_3)_3\}$, 17.0 (C-18), -4.6 {Si(CH₃)₂-}. MASS (FAB): m/z (%): 670 (100, M⁺ + H). Anal. Calcd for (C₄₂H₅₉NO₄Si): C, 75.29; H, 8.88; N, 2.09%. Found: C, 75.45; H, 9.15; N, 2.19%.

N-Methyl amine derivative 15c. As described for 15a, amine 14c (0.2 g, 0.32 mmol), K_2CO_3 (0.22 g, 1.61 mmol), methyl iodide (0.04 ml, 0.64 mmol) in acetone (15 ml) furnished 15c (0.19 g, 93%) as a viscous oil, $R_f = 0.55$ (5% ethyl acetate in hexane), $[a]_D^{20}$ +12 (*c* 2, MeOH). IR (Neat): 3438, 2931, 2361, 1731, 1461, 1378, 1249, 1093, 772 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 7.36–7.23 (m, 5H, –O–CH₂–C₆ H_5), 7.13 (d, 1H, J = 8.7, C-1-H), 6.70 (dd, 1H, $J_I = 8.7$, $J_2 = 2.5$, C-2-H), 6.62 (d, 1H, J = 2.4, C-4-H), 4.95

 $(s, 2H, -O-CH_2-C_6H_5), 3.62(s, 3H, -CO_2CH_3), 3.55-3.42(m, 2H, -CO_2CH_3), 3.55-3.42(m, 2H, -CO_2CH_3))$ C-16-H₂), 2.81-2.75 {m, 3H, -CH-CH-(CH₃)-CH₂-CH₃ and C- $6-H_2$, 2.36–2.34 (m, 2H, C-17- H_2), 2.27 (s, 3H, $-N-CH_3$), 2.26– 2.14 (m, 2H), 1.97-1.10 (m, 3H), 1.84-1.60 (m, 3H), 1.34-1.22 (m, 4H), 0.97-0.90 (m, 1H), 0.84 (bs, 12H, $-C(CH_3)_3$), 0.77-0.73(m, 6H, C-18- H_3), 0.72–0.66 (m, 1H), 0.00 {s, 6H, $-Si(CH_3)_2$ -}. ¹³C NMR (75 MHz, CDCl₃): δ 172.8 (-CO₂CH₃), 156.7 (C-3), 137.9 (C-5), 137.6, 133.1 (C-10), 128.5, 127.8, 127.4, 126.5 (C-1), 114.4 (C-4), 112.8 (C-2), 74.0 {-CH-CH-(CH₃)-CH₂-CH₃}, 69.9 (-O-CH₂-C₆H₅), 67.4 (C-17), 64.4 (C-16), 50.5 (-CO₂CH₃), 44.1, 43.6, 41.8, 40.2 40.2 (C-13), 39.0 (-N-CH₃), 37.7, 34.5 {-CH-CH-(CH₃)–CH₂–CH₃}, 32.8, 30.6 (C-6), 29.6, 27.6 (C-12), 26.5 (C-11), 26.0 $\{-C(CH_3)_3\}, 25.1 \{-CH-CH-(CH_3)-CH_2-CH_3\}, 18.4$ $\{-C(CH_3)_3\}, 16.4 \{-CH-CH-(CH_3)-CH_2-CH_3\}, 16.1 (C-18),$ 11.3 {-CH-CH-(CH₃)-CH₂-CH₃}, -5.1 {Si(CH₃)₂-}. MASS (FAB): m/z (%): 636 (100, M⁺ + H). Anal. Calcd for $(C_{39}H_{61}NO_4Si)$: C, 73.65; H, 9.67; N, 2.20%. Found: C, 73.99; H, 9.79; N, 2.30%.

Alcohol 16a. Compound 15a (0.15 g, 0.25 mmol) was heated at 60 °C in acetic acid (3 ml), water (1 ml), THF (1 ml) (3 : 1 : 1) for 2 h. The solvent was removed under vacuum. The residue was chromatographed over silica gel and elution with 30% ethyl acetate in hexane ($R_f = 0.45$) furnished **16a** (0.08 g, 66%) as a white semi-solid, $[a]_{D}^{20}$ +36.5 (c 2, MeOH). IR (KBr): 3400, 2927, 1732, 1598, 1353, 1236, 1037, 744 $\rm cm^{-1}.$ $^1\rm H$ NMR (200 MHz, CDCl₃): δ 7.40–7.26 (m, 5H, –O–CH₂–C₆H₅), 7.14 (d, 1H, J = 8.6, C-1-H), 6.72 (dd, 1H, $J_1 = 8.6$, $J_2 = 2.4$, C-2-H), 6.64 (d, 1H, J = 2.4, C-4-H), 5.00 (s, 2H, -O-CH₂-C₆H₅), 3.70 (s, 3H, -CO₂CH₃), 3.69–3.35 (m, 3H, C-16-H₂ and -CH-CH₃), 2.90–2.71 (m, 2H, C-6-H₂), 2.31 (s, 3H, -N-CH₃), 2.30-2.11 (m, 2H, C-17- H_2), 2.07–2.03 (m, 2H), 1.75–1.62 (m, 4H), 1.49–1.21 (m, 5H), 1.21 (d, 3H, J = 7.1, $-CH-CH_3$), 0.82 (s, 3H, C-18- H_3). ¹³C NMR (50 MHz, CDCl₃): δ 174.3 (-CO₂CH₃), 157.1 (C-3), 138.2 (C-5), 137.7, 133.2 (C-10), 128.9, 128.2, 127.8, 127.0 (C-1), 114.7 (C-4), 112.9 (C-2), 70.3 (-O-CH₂-C₆H₅), 67.3 (C-17), 64.1 (C-16), 63.7 (-CH-CH₃), 51.6 (-CO₂CH₃), 43.9, 43.5, 42.7, 39.7 (-N-CH₃), 39.3 (C-13), 38.3, 33.0, 31.0 (C-6), 27.8 (C-12), 27.4 (C-11), 18.3 (-CH-CH₃), 15.4 (C-18). MASS (FAB): *m*/*z* (%): 480 (100, M⁺). Anal. Calcd for (C₃₀H₄₁NO₄): C, 75.12; H, 8.62; N, 2.92%. Found: C, 75.38; H, 8.76; N, 3.15%.

Alcohol 16b. As described for 16a, 15b (0.17 g, 0.25 mmol) in AcOH-H₂O-THF (3:1:1) furnished 16b (0.09 g, 63%) as a white semi-solid, $R_{\rm f} = 0.5$ (30% ethyl acetate in hexane), $[a]_{\rm D}^{20} + 16.5$ (c 2, MeOH). IR (KBr): 3980, 2925, 1730, 1606, 1498, 1454, 1222, 1029, 763 cm⁻¹. ¹H NMR (200 MHz, CDCl₃): δ 7.44–7.14 (m, 11H, –O– $CH_2-C_6H_5$, $-CH-CH_2-C_6H_5$ and C-1-H), 6.78 (dd, 1H, $J_1 = 8.6$, $J_2 = 2.4$, C-2-*H*), 6.64 (d, 1H, J = 2.4, C-4-*H*), 5.01 (s, 2H, -O-CH₂-C₆H₅), 3.65 (s, 3H, -CO₂CH₃), 3.57-3.44 (m, 3H, C-16-H₂ and -CH-CH2-C6H5), 3.05-2.85 (m, 2H, -CH-CH2-C6H5), 2.81-2.65 (m, 2H, C-6- H_2), 2.55–2.25 (m, 5H, C-17- H_2 and –N–C H_3), 2.19–1.12 (m, 11H), 0.72 (s, 3H, C-18-H₃). ¹³C NMR (50 MHz, CDCl₃): δ 173.2 (-CO₂CH₃), 157.1 (C-3), 139.0 (C-5), 138.2, 137.7, 133.4 (C-10), 129.9, 128.9, 128.6, 128.2, 127.8, 127.0 (C-1), 126.7, 114.7 (C-4), 112.9 (C-2), 70.9 (-CH-CH₂-C₆H₅), 70.3 (-O-CH₂- C_6H_5), 67.9 (C-17), 64.1 (C-16), 51.5 ($-CO_2CH_3$), 43.6, 43.2, 42.5, 40.2 (-N-CH₃), 40.0 (C-13), 37.6, 36.4 (-CH-CH₂-C₆H₅), 33.1, 31.0 (C-6), 27.8 (C-12), 27.1 (C-11), 17.9 (C-18). MASS (FAB):

m/z (%): 556 (100, M⁺ + H). Anal. Calcd for (C₃₆H₄₅NO₄): C, 77.80; H, 8.16; N, 2.52%. Found: C, 77.99; H, 8.37; N, 2.77%.

Alcohol 16c. As described for 16a, 15c (0.15 g, 0.24 mmol) in AcOH-H₂O-THF (3 : 1 : 1) furnished 16c (0.1 g, 70%) as a transparent viscous liquid, $R_{\rm f} = 0.55$ (30% ethyl acetate in hexane), $[a]_{D}^{20}$ +35 (c 0.8, MeOH). IR (Neat): 3989, 2927, 1734, 1611, 1490, 1459, 1228, 1021, 768 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 7.43–7.19 (m, 6H, –O–CH₂–C₆H₅ and C-1-H), 6.79 (dd, 1H, $J_1 = 8.7$, $J_2 = 2.5$, C-2-H), 6.71 (d, 1H, J = 2.5, C-4-H), 5.03 (s, 2H, $-O-CH_2-C_6H_5$), 3.70 (s, 3H, $-CO_2CH_3$), 3.45–3.39 (m, 2H, C-16-H₂), 2.91-2.83 {m, 3H, {-CH-CH-(CH₃)-CH₂-CH₃ and C-6- H_2 }, 2.48–2.39 (m, 2H, C-17- H_2), 2.35 (s, 3H, –N–C H_3), 2.71–2.27 (m, 2H), 2.03–1.67 (m, 9H), 1.45–1.32 (m, 2H), 0.95–0.8 (m, 11H, C-18- H_3). ¹³C NMR (50 MHz, CDCl₃): δ 172.7 (-CO₂CH₃), 156.2 (C-3), 138.1 (C-5), 137.2, 133.5 (C-10), 128.4, 127.6, 127.5, 126.4 (C-1), 114.7 (C-4), 112.2 (C-2), 74.2 {-CH-CH-(CH₃)-CH₂-CH₃}, 70.0 (-O-CH₂-C₆H₅), 67.0 (C-17), 64.2 (C-16), 50.2 (-CO₂CH₃), 44.5, 43.2, 41.7, 40.5 (C-13), 39.5 (-N-CH₃), 37.5, 34.5 {-CH-CH-(CH₃)-CH₂-CH₃}, 32.2, 30.5 (C-6), 27.5 (C-12), 26.2 (C-11), 25.5 {-CH-CH-(CH₃)-CH₂-CH₃}, 16.5 {-CH-CH-(CH₃)-CH₂CH₃}, 16.2 (C-18), 11.2 {-CH-CH-(CH₃)–CH₂–CH₃}. MASS (FAB): *m*/*z* (%): 507 (100, M⁺).

Hydroxy acid 17a. To a solution of 16a (0.1 g, 0.21 mmol) in dioxane (5 ml) was added 1 N NaOH (0.1 ml) and the resulting solution was heated at 70 °C for 2 h. The reaction mixture was cooled and acidified with 1 N HCl. The resulting solution was extracted with ethyl acetate and dried over Na2SO4 and concentrated in vacuo. The residue was chromatographed over silica gel and elution with 10% methanol in chloroform ($R_{\rm f} = 0.5$) furnished **17a** (0.07 g, 70%) as a light yellow foam, $[a]_{D}^{20}$ +52 (c 2, MeOH). IR (KBr): 3442, 2925, 1707, 1596, 1500, 1352, 1028, 800, 582 cm⁻¹. ¹H NMR (200 MHz, CDCl₃): δ 7.42–7.29 (m, 5H, $-O-CH_2-C_6H_5$, 7.13 (d, 1H, J = 8.6, C-1-H), 6.75 (dd, 1H, $J_1 =$ 8.6, $J_2 = 2.4$, C-2-*H*), 6.67 (d, 1H, J = 2.4, C-4-*H*), 4.99 (s, 2H, $-O-CH_2-C_6H_5$), 3.91–3.84 (m, 2H, C-16- H_2), 3.63–3.53 (m, 1H, $-CH-CH_3$), 3.03–2.95 (m, 2H, C-6- H_2), 2.86 (s, 3H, $-N-CH_3$), 2.82–2.69 (m, 2H, C-17-H₂), 2.30–2.25 (m, 2H), 2.04–1.73 (m, 4H), 1.49 (d, 3H, J = 7, $-CH-CH_3$), 1.48–1.25 (m, 5H), 0.94 (s, 3H, C-18-H₃). ¹³C NMR (50 MHz, CDCl₃): δ 171.5 (-CO₂H), 157.2 (C-3), 138.0 (C-5), 137.6, 132.3 (C-10), 128.9, 128.2, 127.8, 126.9 (C-1), 114.8 (C-4), 113.0 (C-2), 70.3 (-O-CH₂-C₆H₅), 66.5 (C-17), 65.3 (C-16), 62.9 (-CH-CH₃), 43.3, 42.9, 41.1 (-N-CH₃), 38.6 (C-13), 37.8, 32.5, 30.8 (C-6), 27.5 (C-12), 27.0 (C-11), 18.8 (-CH-CH₃), 12.1 (C-18). MASS (FAB): *m*/*z* (%): 466 (100, M⁺). Anal. Calcd for (C₂₉H₃₉NO₄): C, 74.81; H, 8.44; N, 3.01%. Found: C, 74.99; H, 8.72; N, 3.11%.

Hydroxy acid 17b. As described for **17a**, **16b** (0.10 g, 0.18 mmol) in dioxane (5 ml), 1 N NaOH (0.1 ml) furnished **17b** (0.07 g, 67%) as a light yellow foam, $R_{\rm f} = 0.55$ (10% methanol in chloroform), $[a]_{\rm D}^{20}$ +49 (*c* 2, MeOH). IR (KBr): 3445, 2923, 1709, 1593, 1506, 1354, 1026, 801, 584 cm⁻¹. ¹H NMR (200 MHz, CDCl₃): δ 7.41–7.12 (m, 10H, –O–CH₂–C₆H₅, –CH–CH₂–C₆H₅) 7.03 (d, 1H, J = 8.6, C-1-*H*), 6.67 (dd, 1H, $J_I = 8.6$, $J_2 = 2.4$, C-2-*H*), 6.57 (d, 1H, J = 2.4, C-4-*H*), 5.01 (s, 2H, –O–CH₂–C₆H₅), 3.95–3.42 (m, 1H, –CH–CH₂–C₆H₅), 3.75–3.61 (m, 2H, C-16-H₂), 3.43–2.29 (m, 2H, –CH–CH₂–C₆H₅), 2.98–3.18 (m, 2H, C-6-H₂), 2.91 (s, 3H, –N–CH₃), 2.74–2.67 (m, 2H, C-17-H₂), 2.18–1.13 (m,

11H), 0.75 (s, 3H, C-18- H_3). ¹³C NMR (75 MHz, CDCl₃): δ 171.4 (- CO_2 H), 156.8 (C-3), 137.9 (C-5), 137.6, 137.0, 132.2 (C-10), 129.5, 128.7, 128.5, 127.8, 127.4, 126.8 (C-1), 126.5, 114.4 (C-4), 112.6 (C-2), 71.9 (-CH-CH₂-C₆H₅), 69.9 (-O-CH₂-C₆H₅), 66.2 (C-17), 62.7 (C-16), 42.9, 42.7, 42.3, 41.3 (N-CH₃), 38.8 (C-13), 37.4, 33.6 (-CH-CH₂-C₆H₅), 32.1, 30.4 (C-6), 27.1 (C-12), 26.5 (C-11), 17.8 (C-18). MASS (FAB): m/z (%): 542 (100, M⁺ + H). Anal. Calcd for (C₃₅H₄₃NO₄): C, 77.60; H, 8.00; N, 2.59%. Found: C, 77.86; H, 8.26; N, 2.78%.

Hydroxy acid 17c. As described for **17a**, **16c** (0.1 g, 0.19 mmol) in dioxane (5 ml), 1 N NaOH (0.1 ml) furnished 17c (0.07 g, 72%) as a light yellow viscous oil, $R_{\rm f} = 0.6$ (10% methanol in chloroform), $[a]_{D}^{20}$ +28 (c 2, MeOH). IR (Neat): 3438, 2929, 1708, 1587, 1509, 1358, 1029, 805, 589 cm⁻¹. ¹H NMR (200 MHz, CDCl₃): δ 7.38-7.11 (m, 6H, -O-CH₂-C₆H₅, and C-1-H), 6.77-6.69 (m, 2H, C-2-H and C-4-H), 4.99 (s, 2H, -O-CH₂-C₆H₅), 3.77-3.73 (m, 2H, C-16-H₂), 2.80-2.73 {m, 3H, -CH-CH-(CH₃)-CH₂-CH₃ and C- $6-H_2$, 2.38–2.26 (m, 2H, C-17- H_2), 2.23 (s, 3H, $-N-CH_3$), 2.16– 1.39 (m, 13H), 1.00–0.89 (m, 8H), 0.59 (s, 3H, C-18-H₃). ¹³C NMR (50 MHz, CDCl₃): δ 171.5 (-CO₂H), 157.2 (C-3), 138.1 (C-5), 137.9, 132.5 (C-10), 128.9, 128.2, 127.8, 126.9 (C-1), 114.9 (C-4), 112.9 (C-2), 73.5 {-CH-CH-(CH₃)-CH₂-CH₃}, 70.2 (-O-CH₂-C₆H₅), 66.9 (C-17), 62.4 (C-16), 44.6, 43.3, 42.4, 42.0 (C-13), 38.4 (-N-CH₃), 36.4, 34.9 {-CH-CH-(CH₃)-CH₂-CH₃}, 32.6, 30.7 (C-6), 27.5 (C-12), 26.6 (C-11), 25.9 {-CH-CH-(CH₃)-CH₂-CH₃}, 16.9 {-CH-CH-(CH₃)-CH₂-CH₃}, 14.6 (C-18), 12.4 $\{-CH-CH-(CH_3)-CH_2-CH_3\}$. MASS (FAB): m/z (%): 507 (100, M⁺).

Lactone 3a. A stirred solution of hydroxy acid 17a (20 mg, 0.04 mmol) in THF (1 ml) at RT under N₂ was treated with triethyl amine (0.06 ml, 0.43 mmol) followed by 2,4,6-trichlorobenzoyl chloride (0.04 ml, 0.26 mmol) and it was stirred overnight. The solution was diluted with toluene (5 ml) and added dropwise over 4 h to a refluxing solution of DMAP (60 mg, 0.51 mmol) in toluene (30 ml) under N₂. The reaction was further stirred for 2 h at reflux, and allowed to cool to room temperature. It was diluted with ethyl acetate and washed with a saturated solution of NaHCO₃. The layers were separated and the water layer was extracted with ethyl acetate. The combined organic extracts were washed with 0.1 M aq. HCl, brine, dried over Na₂SO₄ and concentrated under vacuum. The residue was chromatographed over silica gel and elution with 10% ethyl acetate in hexane ($R_{\rm f} = 0.4$) furnished **3a** (10 mg, 53%) as a light yellow oil. ¹H NMR (300 MHz, CDCl₃): δ 7.43–7.31 (m, 5H, $-O-CH_2-C_6H_5$), 7.21 (d, 1H, J = 8.6, C-1-H), 6.78 (dd, 1H, $J_1 = 8.6$, $J_2 = 2.4$, C-2-H), 6.71 (d, 1H, J = 2.4, C-4-*H*), 5.03 (s, 2H, $-O-CH_2-C_6H_5$), 4.79–4.71 (m, 1H, C-16-*H*), 3.98-3.81 (m, 1H, C-16-H), 3.62-3.60 (m, 1H, -CH-CH₃), 2.88-2.78 (m, 2H, C-6- H_2), 2.55 (s, 3H, $-N-CH_3$), 2.48–2.20 (m, 2H, C-17-H₂), 2.00–1.97 (m, 2H), 1.84–1.73 (m, 3H), 1.43–1.30 (m, 3H), 1.16 (d, 3H, J = 6.6, $-CH-CH_3$), 0.88–0.81 (m, 3H), 0.61 (s, 3H, C-18- H_3). MASS (FAB): m/z (%): 448 (100, M⁺ + H). Exact Mass (ESI-MS) calculated for C₂₉H₃₇NO₃ [M]⁺: 447.27735, found: 447.27740.

Lactone 3b. As described for **3a**, **17b** (20 mg, 0.04 mmol) in THF (1 ml), triethyl amine (0.05 ml, 0.37 mmol), 2,4,6-trichlorobenzoyl chloride (0.03 ml, 0.22 mmol), DMAP (50 mg, 0.44 mmol) in toluene (30 ml) furnished **3b** (12 mg, 63%) as a light

yellow oil, $R_{\rm f} = 0.5$ (10% ethyl acetate in hexane), $[a]_{\rm D}^{20}$ +15.5 (c 2, MeOH). ¹H NMR (300 MHz, CDCl₃): δ 7.45–7.20 (m, 11H, $-O-CH_2-C_6H_5$, $-CH-CH_2-C_6H_5$ and C-1-H), 6.79 (dd, 1H, $J_1 =$ 8.6, $J_2 = 2.4$, C-2-*H*), 6.72 (d, 1H, J = 2.4, C-4-*H*), 5.04 (s, 2H, $-O-CH_2-C_6H_5$, 4.53–4.50 (m, 1H, C-16-H), 4.19–4.05 (m, 1H, C-16-*H*), 3.66–3.64 (m, 1H, –C*H*–CH₂–C₆H₅), 3.07–2.91 (m, 2H, -CH-CH₂-C₆H₅), 2.87-2.86 (m, 2H, C-6-H₂), 2.66 (s, 3H, -N- CH_3 , 2.39–2.27 (m, 2H, C-17- H_2), 2.05–1.81 (m, 4H), 1.68–1.27 (m, 7H), 0.68 (s, 3H, C-18- H_3). ¹³C NMR (50 MHz, CDCl₃): δ 173.9 (-CO-), 156.8 (C-3), 138.9 (C-5), 137.9, 137.3, 133.2 (C-10), 129.3, 128.5, 128.4, 127.8, 127.4, 126.2(C-1), 126.1, 114.4 (C-4), 112.4 (C-2), 71.8($-CH-CH_2-C_6H_5$), 69.9 ($-O-CH_2-C_6H_5$), 67.1 (C-16), 61.1 (C-17), 46.2 (-N-CH₃), 43.1, 42.4, 41.1, 40.4 (C-13), 37.3, 30.4 (C-6), 29.7, 28.6 (-CH-CH2-C6H5), 27.6 (C-12), 26.0 (C-11), 17.9 (C-18). MASS (FAB): *m*/*z* (%): 524 (100, M⁺ + H). Anal. Calcd for (C₃₅H₄₁NO₃): C, 80.27; H, 7.89; N, 2.67%. Found: C, 80.48; H, 7.99; N, 2.86%.

Lactone 3c. As described for 3a, 17c (20 mg, 0.04 mmol) in THF (1 ml), triethyl amine (0.05 ml, 0.39 mmol), 2,4,6-trichlorobenzoyl chloride (0.04 ml, 0.24 mmol), DMAP (60 mg, 0.47 mmol) in toluene (30 ml) furnished 3c (8 mg, 42%) as a light yellow oil, $R_{\rm f} = 0.45$ (10% ethyl acetate in hexane). ¹H NMR (300 MHz, CDCl₃): δ 7.43–7.30 (m, 5H, –O–CH₂–C₆H₅), 7.19 (d, 1H, J = 8.7, C-1-H), 6.77 (dd, 1H, $J_1 = 8.7$, $J_2 = 2.7$, C-2-H), 6.71 (d, 1H, J = 2.7, C-4-H), 5.02 (s, 2H, –O–CH₂–C₆H₅), 4.40–4.21 (m, 1H, C-16-H), 4.21–4.16 (m, 1H, C-16-H), 3.15–3.19 (m, 1H, –CH–CH–(CH₃)–CH₂–CH₃}, 2.86–2.85 (m, 2H, C-6- H_2), 2.80–2.75 (m, 1H), 2.31–2.23 (m, 4H, –N–CH₃), 1.95–1.75 (m, 5H), 1.73–1.07 (m, 7H), 0.94–0.68 (m, 8H), 0.68 (s, 3H, C-18- H_3). MASS (FAB): m/z (%): 490 (100, M⁺ + H).

Acknowledgements

Financial support of this work by DST (SR/FTP/CSA-05/2002) and MOH, New Delhi and a fellowship to Shagufta by CSIR are gratefully acknowledged. Dr Raja Roy is sincerely thanked for recording ¹H-¹H COSY, HSQC and HMBC spectra.

References

- (a) H. Floss, Nat. Prod. Rep., 1997, 14, 433–452; (b) K. C. Nicolaou, D. Vourloumis, N. Winssinger and P. S. Baran, Angew. Chem., Int. Ed., 2000, 39, 44–122 and references cited therein.
- 2 (a) G. Mehta and V. Singh, Chem. Soc. Rev., 2002, 31, 324-334 and references cited therein; (b) L. F. Tietze, R. Hannemann, W. Buhr, M. Logers, P. Menningen, M. Lieb, D. Starck, T. Grote, A. Doring and I. Schuberth, Angew. Chem., Int. Ed. Engl., 1996, 35, 2674-2676; (c) L. F. Tietze, G. Schneider, J. Wolfling, A. Fecher, T. Nobel, S. Peterson, I. Schuberth and C. Wulff, Chem.-Eur. J., 2000, 6, 3755-3760; (d) F. De Riccardis, I. Izzo, M. Di Filippo, G. Sodano, F. D'Acquisto and R. Carnuccio, Tetrahedron, 1997, 53, 10871-10882; (e) J. J. Masters, D. K. Jung, S. J. Danishefsky, L. B. Zinder, T. K. Park, R. C. A. Isaccs, C. A. Alaimo and W. B. Young, Angew. Chem., Int. Ed. Engl., 1995, 34, 452–455; (f) J. Wang and P. J. De Clercq, Angew. Chem., Int. Ed. Engl., 1995, 34, 1749-1752; (g) G. B. Jones, G. Hynd, J. M. Wright, A. Purohit, G. W. Plourde, II, R. S. Huber, J. E. Mathews, A. Li, M. W. Kilgore, G. J. Bubley, M. Yanacisin and M. A. Brown, J. Org. Chem., 2001, 66, 3688-3695; (h) V. Singh, S. Lahiri, V. V. Kane, T. Stey and D. Stalke, Org. Lett., 2003, 5, 2199-2202; (i) A. Sadownik, G. Deng, V. Janout and S. L. Regen, J. Am. Chem. Soc., 1995, 117, 6138-6139; (j) G. Mehta, S. Muthusamy, B. G. Maiya and M. Sirish, J. Chem. Soc., Perkin Trans. 1, 1996, 2421-2423; (k) S. D. Kuduk, F. F. Zheng, L. S. Lorenzino, N. Rosen and S. J. Danishefsky, Bioorg. Med. Chem. Lett.,

1999, **9**, 1233–1238; (*l*) E. Alvaro, M. C. de la Torre and M. A. Sierra, *Chem. Commun.*, 2006, **9**, 985–987; (*m*) J. F. Billing and U. J. Nilsson, *J. Org. Chem.*, 2005, **70**, 4847–4850.

- 3 M. Mokotoff, M. Zhao, R. J. Marshall, E. Winslow, L. K. Wong and Q.-J. Liao, *Steroids*, 1990, **55**, 399–404.
- 4 L. Lapatsanls, J. Chem. Eng. Data, 1980, 25, 287-289.
- 5 C. Wang, M. Zhao, J. Yang and S. Peng, Steroids, 2001, 66, 811-815.
- 6 F. Kong and R. J. Andersen, J. Org. Chem., 1993, 58, 6924–6927.
- 7 A. X. Yan, R. Y. K. Chan, W. S. Lau, K. S. Lee, M. S. Wong, G. W. Xing, G. I. Tian and Y. H. Ye, *Tetrahedron*, 2005, **61**, 5933– 5941.
- 8 T. Matsumoto, M. Watanabe, S. Mataka and T. Thiemann, *Steroids*, 2003, **68**, 751–757.

- 9 G. R. Fluoret, W. Cole and U. Biermacher, J. Med. Chem., 1972, 15, 1281–1283.
- 10 L. F. Tietze, K. M. Sommer, G. Schneider, P. Tapolesanyi, J. Wolfling, P. Muller, M. Noltemeyer and H. Terlau, *Synlett*, 2003, 1494– 1496.
- 11 D. S. Fischer, L. W. L. Woo, M. F. Mahon, A. Purohit, M. J. Reed and B. V. L. Potter, *Bioorg. Med. Chem.*, 2003, **11**, 1685– 1700.
- 12 N. S. Leeds, D. K. Fukushima and T. F. Gallagher, J. Am. Chem. Soc., 1954, 76, 2943–2948.
- 13 J. K. Mishra and G. Panda, Synthesis, 2005, 1881-1887.
- 14 J. Inanaga, K. Hirata, H. Saeki, T. Katsuki and M. Yamaguchi, Bull. Chem. Soc. Jpn., 1979, 52, 1989–1993.